

Example Report

Prepared for:

Customer name:

Internal project number: Quote number:

Version: 1 Date: 21-Feb-22





Goal

In this study, 1 transgenic CHO cell line with the vector xxxxx sequence was analyzed. The aim of this analysis was to:

- 1. Study the vector integrity;
 - 1) Determine the presence of sequence variants and their allele frequency.
 - 2) Determine the presence of vector-vector breakpoints that represent concatemers of multiple copies of the vector and/or structural rearrangements in a single vector sequence.
- 2. Identify vector integration site(s) and breakpoint sequences between the vector and genome.
- 3. Assess the presence of structural variants surrounding the vector integration site(s).
- 4. Estimate the copy number of the vector.

An overview of the TLA technology and technical details of the performed analyses is provided in the manual "Introduction to the terminology and methods used in TLA analyses v2".

Summary

Sample	Vector Integrity	Integration site(s)	Structural variants at the integration site	Copy number estimation
Sample 1	6 sequence variants, 2 structural variants	Chr3: 169,680,259 - 169,680,260	no	3-5

Conclusion

In Sample 1, 3-5 copies of the vector have integrated in chr 3. 6 sequence variants and 2 structural variants are found within the integrated vector sequence.



Methods

TLA, sequencing and data mapping

Viable frozen CHO-K1 cells were used and processed according to Cergentis' TLA protocol (de Vree et al. Nat Biotechnol. Oct 2014). An overview of the TLA technology and technical details of the performed analyses is provided in the manual "Introduction to the terminology and methods used in TLA analyses v1".

TLA was performed with 2 independent primer sets specific for the vector sequence (Table 1).

Table 1: Primers used in TLA analysis

Primer set	Name/Viewpoint	Direction	Binding position	Sequence
1	Amp	Rv	132	Х
		Fw	256	Х
2	GOI	Rv	2,745	Х
		Fw	3,186	Х

The NGS reads were aligned to the vector sequence and host genome. The Chinese Hamster CriGri-PICRH1.0 genome assembly GCF_003668045.3 was used as host reference genome sequence.



Results Sample 1

Vector integrity

Figure 1 depicts the NGS coverage across the vector sequence using primer set 1. Same results were obtained with primer set 2.



Figure 1: NGS sequencing coverage across the vector with primer set 1. The black arrow indicates the primer location. The green arrows indicate the locations of the identified vector-genome breakpoint sequences (described below). Y-axis is limited to 1000x. In an actual report the data of all primer sets will be presented.

High coverage is observed across the complete vector sequence Vector: 1-10,593. Local dips in coverage are due to GC rich regions that are less efficiently sequenced.

Sequence variants and structural variants were called in the covered regions.

Sequence variants

Detected sequence variants are presented in table 2. A total of 6 sequence variants were identified in the sample. Sequence variants at or near 100% mutation frequency most likely represent deviations present in the provided reference sequence of the vector before its introduction into the sample.

Table 2: Identifi	ied sequer	nce variants
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				Primer set 1		Primer set 2	
Region	Position	Ref	Mut	Cov	%	Cov	%
Amp	141	А	С	21,254	20	788	25
GOI	1,013	Т	-4AGTT	1,881	100	7,501	100
GOI	2,956	Т	С	1,278	27	34,122	21
GOI	5,698	А	+1G	751	18	2,221	15
Backbone	9,487	Т	G	1,358	100	1,523	99
Backbone	10,037	G	А	2,145	20	854	20

'+' indicates an insertion; '-' indicates a deletion.

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Vector concatemerization and structural variants

The identified vector-vector breakpoint sites are shown in table 3. In the accompanying excel tables the sequences and frequencies of the breakpoints are presented. In total, 2 structural variants were identified. Intact reads were also found at all positions indicating that (partial) vector sequences have concatemerized.

Table 3: Vector-vector breakpoints

Breakpoint	Vector		Vector		Orientation of the breakpoint	Homology	Insert
1	→	6,945	7,657	→	tail to head	1	-
2	→	14	5,051	÷	tail to tail	-	2



Integration sites

Whole genome coverage plot



Figure 2: TLA sequence coverage across the Chinese Hamster genome using primer set 1. The chromosomes are indicated on the y-axis, the chromosomal position on the x-axis. Identified integration site is encircled in green.

As shown in figure 2, the vector has integrated on chromosome 3. Similar results were obtained with primer set 2.



Locus-wi	ide coverage
	•
Set 1	ne n
Gene	I I

Figure 3: TLA sequence coverage (in blue) across the vector integration locus, chr3:169,530,000-169,830,000. The green arrow indicates the location of the breakpoint sequences. Y-axis is limited to 200x. In an actual report the data of all primer sets will be presented.

Coverage is observed across the vector integration site as shown in figure 3.

Breakpoint sequences

The following breakpoint sequences were identified marking the vector integration:

5' integration site:

The coverage profile in figure 3 shows that no genomic rearrangements have occurred in the region of the integration site.

From this data it is concluded that the vector has integrated chr3: 169,680,259 - 169,680,260. According to Refseq, there are no genes annotated here.





Copy number estimation

In Sample 1, the coverage on the vector-side is 4-5 times higher than on the genome-side of the integration site. 1 integration site and 2 vector-vector breakpoints are found. The copy number is estimated to be 3-5 copies.

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QC information

Sample and Study details

Sample receipt date Condition of sample at receipt Start date in the lab Sequencing run Deviations from the protocol TLApp version

DNA was received frozen in a Thermo Scientific Matrix 96 tube rack, Labeled gDNA xxxxx

Study Personnel Lab technician Lab technician qPCR Data Analyst QC Analysis and Report



Scientific-approval Date Signature

Quality control

The results are independently verified and reviewed and are an accurate and complete representation of the study. TLA processing of cells, NGS sequencing, and data analysis (except for copy number) are ISO/IEC 17025:2017 accredited by the Dutch Accreditation Council RvA, Registration number L671.

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